

ORIGINAL ARTICLE

Sheng-Yang Wang · Jyh-Horng Wu · Sen-Sung Cheng  
Chiu-Ping Lo · Hsing-Ning Chang · Lie-Fen Shyur  
Shang-Tzen Chang

## Antioxidant activity of extracts from *Calocedrus formosana* leaf, bark, and heartwood

Received: December 9, 2002 / Accepted: September 8, 2003

**Abstract** *Calocedrus formosana* Florin (Cupressaceae) is an endemic tree to Taiwan. To evaluate the antioxidant activity of the plant extracts from heartwood, bark, and leaf of *C. formosana*, assays for 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical and superoxide anion scavenging activities, as well as prevention of DNA strand cleavage were performed in this study. Similar IC<sub>50</sub> values against the DPPH radical were found for the heartwood and bark extracts at approximately 23 µg/ml. Moreover, the heartwood extract exhibited the highest inhibitory activity against superoxide radicals among the test samples; a 2.3-fold lower value of IC<sub>50</sub> for superoxide radical inhibition was found in the heartwood extract relative to that of (+)-catechin. Much less effect on inhibition of DPPH and superoxide radicals was found from the leaf extract of *C. formosana*. More than 70% of superoxide radicals were inhibited in the presence of 10 µg/ml heartwood extract, whereas only 15% inhibition was obtained from the leaf extract. The heartwood extract, at a dose of approximately 0.5 mg/ml, apparently completely prevented the ΦX174 supercoiled DNA cleavage induced by ultraviolet photolysis of H<sub>2</sub>O<sub>2</sub>, as judged by agarose gel electrophoresis. This report also suggests that the antioxidant activities of the plant extracts of *C. formosana* are in good correlation with their phenolic contents.

**Key words** *Calocedrus formosana* · Heartwood extract · Antioxidant activity · Total phenolic content

S.-T. Chang (✉) · J.-H. Wu · S.-S. Cheng  
Department of Forestry, National Taiwan University, No. 1 Section 4  
Roosevelt Road, Taipei 106, Taiwan  
Tel. +886-2-23630231-3196; Fax +886-2-23654520  
e-mail: peter@ntu.edu.tw

L.-F. Shyur · S.-Y. Wang · C.-P. Lo · H.-N. Chang  
Institute of BioAgricultural Sciences, Academia Sinica, Taipei 115,  
Taiwan  
Tel. +886-2-27899322; Fax +886-2-27899322  
e-mail: lfshyur@ccvax.sinica.edu.tw

### Introduction

Secondary metabolites of trees commonly play important roles in specific chemical or physical properties of the wood, such as color, odor, durability, and others. Furthermore, many natural compounds isolated from woody plants have been found to have significant pharmacological activities, with great potential for further development as drugs or drug analogs for human disease control. Taxol and podophyllotoxin isolated from conifers are two typical and well-known examples, which have been recognized as potent antitumor drugs. Reactive oxygen species (ROS), e.g., superoxide radicals, hydroxyl radicals, and hydrogen peroxide, have been proposed as significant causative factors in some radical-mediated conditions including aging,<sup>1</sup> cancer,<sup>2,3</sup> and cardiovascular disease.<sup>4</sup> The search for potent natural antioxidants, especially from plant sources, as nutritional supplements, health food, and/or phytomedicine has become an important research issue at a world-wide level. Many physicians and researchers now contemplate the use of antioxidant treatments as a key strategy for inhibiting or reversing the process of carcinogenesis.<sup>5</sup>

*Calocedrus formosana* Florin (Cupressaceae) is an endemic tree that grows at elevations of 800–1500 m in Taiwan's central mountains. The physico-chemical properties of the timbers of this plant are well recognized for their decay resistance and excellent durability. With regard to the phytochemical studies of *C. formosana*, more than 30 compounds have been isolated from the plant including monoterpenes, diterpenes, lignans, and steroids.<sup>6–10</sup> Recently, we identified that the major compound of the exudates from the bark surface of *C. formosana* is ferruginol.<sup>11</sup> However, we have found nothing regarding the potential human health benefits of the extracts and phytocompounds of this plant. In our recent studies on *Taiwania cryptomerioides* (Taxodiaceae), another indigenous conifer tree species grown in Taiwan, we identified several forms of diterpenes from its heartwood, and found that ferruginol exhibited the most significant antioxidant activity among the diterpenes.<sup>12</sup> Based on the characteristics of the compounds isolated

from *C. formosana* and to explore the additional utilization value of extractives from *C. formosana*, we found it of interest to investigate the antioxidant activity of this plant.

In this study a number of in vitro assays including 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical scavenging activity test, nitroblue tetrazolium chloride (NBT) assay, and hydroxyl radical-induced DNA strand scission assay were performed to evaluate the antioxidant activities of *C. formosana* extracts. Several previous reports have demonstrated that phenolic compounds have multiple biological effects, especially for antioxidant activity.<sup>13</sup> Therefore, we quantitatively analyzed the phenolics in *C. formosana* and compared their distributions in its various tissues. Our results demonstrated that the total ethanolic extract of heartwood contains the highest amount of phenolic compounds, and that it also exhibited the most significant antioxidant activity among the tested extracts. Moreover, the inhibition of heartwood extract for ROS is comparable to that of (+)-catechin, which is well known for potential antioxidant activity. This report indicates that plant extracts or derived compounds from *C. formosana* may serve as good candidates for further development as nutraceutical or antioxidant remedies.

## Materials and methods

### Preparation of plant extracts

Leaf, bark and heartwood of *Calocedrus formosana* were collected from the experimental forest of National Taiwan University in central Taiwan. The dried samples were cut into small pieces and soaked in 70% (v/v) ethanolic aqueous solution at room temperature for 7 days. The extract was decanted, filtered under vacuum, concentrated in a rotary evaporator, and then lyophilized. The resulting powder extracts were employed for the current study.

### Reagents

1,1-Diphenyl-2-picrylhydrazyl (DPPH), hypoxanthine, xanthine oxidase, nitroblue tetrazolium chloride (NBT),  $\Phi$ X174 RF1 supercoiled DNA, Folin-Ciocalteu's reagent, (+)-catechin, and gallic acid were purchased from Sigma (St. Louis, MO, USA). The other chemicals and solvents used in this study were of the highest quality available.

### DPPH assay

Scavenging actions of DPPH free radical by *C. formosana* extracts were measured as follows. The reaction mixture contained 1000  $\mu$ l of 0.1 mM DPPH-ethanol solution, 450  $\mu$ l of 0.05 M Tris-HCl buffer (pH 7.4), and 50  $\mu$ l of test samples (final concentrations were 1, 5, 10, 30, and 50  $\mu$ g/ml, respectively) or methanol (control). Reduction of the DPPH free radical was measured by reading the absorbance at 517 nm exactly 30 min after adding each of the extracts.<sup>14</sup>

(+)-Catechin was used as a positive reference in this experiment. The inhibition ratio was expressed as a percentage after being calculated from the following equation: % Inhibition = [(absorbance of control – absorbance of test sample)/absorbance of control]  $\times$  100.

### NBT (superoxide scavenging) assay

Measurement of superoxide radical scavenging activity was carried out based on the method described by Kirby and Schmidt<sup>15</sup> with slight modifications. Twenty microliters of 15 mM Na<sub>2</sub>EDTA in buffer (50 mM KH<sub>2</sub>PO<sub>4</sub>/KOH, pH 7.4), 50  $\mu$ l of 0.6 mM NBT in buffer, 30  $\mu$ l of 3 mM hypoxanthine in 50 mM KOH, 5  $\mu$ l of test samples in methanol (final concentrations were 1, 5, 10, and 50  $\mu$ g/ml, respectively), and 145  $\mu$ l of buffer were mixed in 96-well microplates (Falcon, USA). The reaction was started by adding 50  $\mu$ l of xanthine oxidase solution (1 unit in 10 ml buffer) to the mixture, which was incubated at 25°C. The absorbance at 570 nm was taken every 20 s up to 5 min using a plate reader (Labsystems Multiskan, Finland). The control contained 5  $\mu$ l methanol instead of the sample solution. (+)-Catechin was used as a positive reference. The inhibition ratio was expressed as a percentage after being calculated from the following equation: % Inhibition = [(rate of control – rate of sample reaction)/ rate of control]  $\times$  100.

### Inhibition of hydroxyl radical-induced DNA strand scission

The assay was done according to the method of Keum et al.<sup>16</sup> with minor modifications. The reaction mixture (30  $\mu$ l) contained 10 mM Tris-HCl in 1 mM EDTA buffer (pH 8.0),  $\Phi$ X174 RF1 DNA (0.3  $\mu$ g), and H<sub>2</sub>O<sub>2</sub> (0.04 M). Various amounts of the test extract samples dissolved in 10  $\mu$ l of ethanol (final concentrations of the plant extract in each assay were 1, 10, 100, 500, and 1000  $\mu$ g/ml, respectively) were added prior to the H<sub>2</sub>O<sub>2</sub> addition. Hydroxyl radicals were generated by irradiation of the reaction mixtures at a distance of 5 cm with a 12-W ultraviolet (UV) lamp (Spectroline, Spectronics, USA). After incubation at room temperature for 20 min, the reaction was terminated by the addition of a loading buffer (0.25% bromophenol blue tracking dye and 40% sucrose), and the mixtures were then analyzed by 0.8% submarine agarose gel electrophoresis (70 eV, 1 h). The gels were stained with ethidium bromide, destained in water, and photographed on a transilluminator.

### Determination of total phenolic content in plant extracts

The phenolic contents in the plant extracts of *C. formosana* were measured according to the Folin-Ciocalteu method with slight modification.<sup>17</sup> One milliliter of extract solution in methanol was mixed with an equal volume of 1N Folin-Ciocalteu reagent and allowed to stand for 5 min at room temperature. After addition of 2.0 ml of 20% Na<sub>2</sub>CO<sub>3</sub> and

incubation at room temperature for another 10 min, the mixture was centrifuged and the absorbance of the supernatant was measured at 730 nm on an UV/VIS spectrophotometer. The total phenolic content was expressed as gallic acid equivalents (GAE) in milligrams per gram dry material (plant extract).

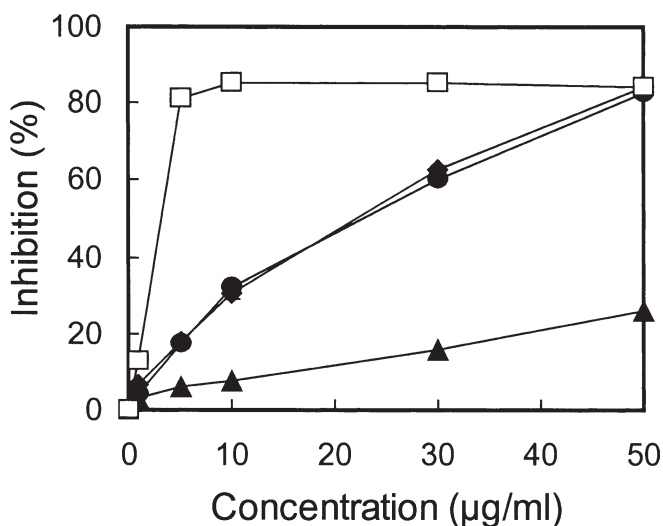
### Statistical analyses

All results were obtained from three independent experiments and expressed as mean  $\pm$  SD. Analyses of variance were performed by ANOVA procedures. Significant differences ( $P < 0.05$ ) were determined by Duncan's multiple-range test.

## Results

### Free radical scavenging activity of plant extracts from *Calocedrus formosana*

The free radical scavenging activities of *Calocedrus formosana* extracts were assessed by DPPH assay. As shown in Fig. 1, both bark and heartwood ethanolic extracts from *C. formosana* exhibited a significant inhibitory activity against the DPPH radical. Similar  $IC_{50}$  values for DPPH radical in bark and heartwood extracts were observed at approximately  $23 \mu\text{g/ml}$ . At  $50 \mu\text{g/ml}$ , two plant extracts inhibited more than 80% of the DPPH radical. Much less activity against the DPPH radical was found from leaf extract as compared with the bark and heartwood extracts (Fig. 1). The  $IC_{50}$  value of a well-known antioxidant compound, (+)-catechin, used as a reference in this study, is approximately  $5 \mu\text{g/ml}$ .



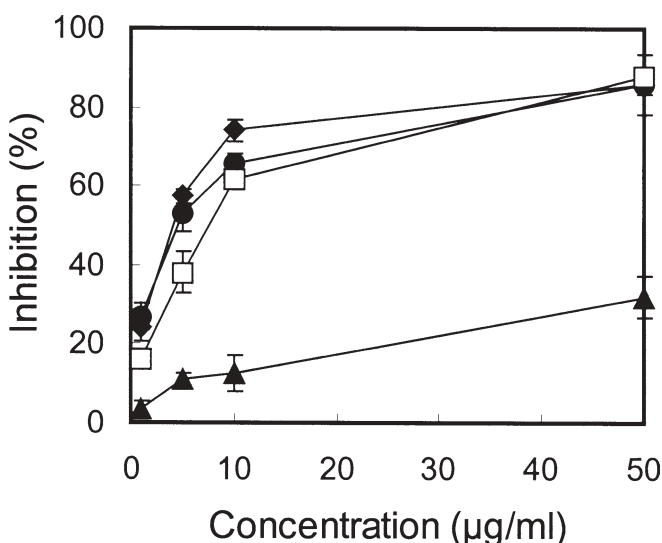
**Fig. 1.** Free-radical scavenging activity of plant extracts from *Calocedrus formosana* measured using the 1,1-diphenyl-2-picrylhydrazyl (DPPH) assay. Results are mean  $\pm$  SD ( $n = 3$ ). Diamonds, heartwood extract; circles, bark extract; triangles, leaf extract; squares, (+)-catechin

### Superoxide scavenging activity of extracts from *Calocedrus formosana*

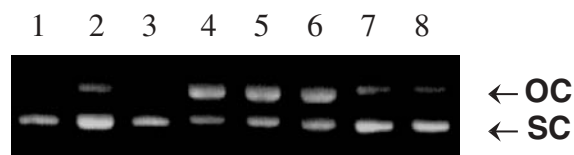
Figure 2 shows the superoxide scavenging activity of the *C. formosana* extracts and (+)-catechin. The heartwood extract shows a more inhibitory activity against superoxide radicals induced by hypoxanthine-xanthine oxidase than the bark and leaf extracts, as well as (+)-catechin at all tested concentrations. The  $IC_{50}$  values for (+)-catechin and the heartwood, bark, and leaf extracts of *C. formosana* were 7, 3, 4, and  $>50 \mu\text{g/ml}$ , respectively (Fig. 2). Even with dosages as low as  $10 \mu\text{g/ml}$ , the heartwood extract was capable of scavenging more than 70% of the generated superoxide radicals in an assay, whereas bark extract, leaf extract, and (+)-catechin scavenged approximately 65%, 15%, and 62%, respectively. These results demonstrate that the ethanolic extracts of *C. formosana* heartwood show a better effect against superoxide radicals as compared with the well-known antioxidant (+)-catechin.

### Inhibition of the hydroxyl radical-induced DNA strand scission

To further characterize the antioxidant bioactivity of the heartwood extract of *C. formosana*, an in vitro assay for determining the prevention of DNA strand cleavage induced by UV photolysis of  $\text{H}_2\text{O}_2$  was performed in this study. Figure 3 shows that the heartwood extract effectively decreases the DNA strand scissions induced by hydroxyl radicals in a dosage-dependent manner. The native  $\Phi\text{X174}$  RF1 DNA was shown with dominant supercoiled form DNA and there was little or no detectable change observed when the DNA was treated with UV or  $\text{H}_2\text{O}_2$  alone, as judged by agarose gel electrophoresis (lanes 1–3).



**Fig. 2.** Superoxide scavenging activity of *C. formosana* extracts measured using the nitroblue tetrazolium chloride (NBT) assay. Results are mean  $\pm$  SD ( $n = 3$ ). Diamonds, heartwood extract; circles, bark extract; triangles, leaf extract; squares, (+)-catechin



**Fig. 3.** Protection effect of heartwood extract of *C. formosana* on DNA strand scission induced by H<sub>2</sub>O<sub>2</sub> and ultraviolet light (UV). Lane 1 represents native  $\Phi$ X174 DNA without any treatment.  $\Phi$ X174 RF1 supercoiled DNA was exposed to UV alone (lane 2), H<sub>2</sub>O<sub>2</sub> alone (lane 3), UV plus H<sub>2</sub>O<sub>2</sub> (lane 4). Lanes 5 to 8 represent supercoiled DNA that was incubated with 50, 100, 500, and 1000  $\mu$ g/ml of the heartwood extracts, respectively, and with H<sub>2</sub>O<sub>2</sub> and UV illumination. DNA was analyzed by 0.8% agarose gel electrophoresis and visualized under the UV illuminator. Arrows indicate distinct forms of the bacteriophage DNA: OC, open circular; SC, supercoiled

**Table 1.** Total phenolic content of plant extracts from *Calocedrus formosana*

Extracts	Total phenolics (mg of GAE/g) <sup>a</sup>
Heartwood	159.5 $\pm$ 1.9
Bark	115.3 $\pm$ 2.3
Leaf	53.5 $\pm$ 1.5

The experiments were measured by Folin–Ciocalteu method as described in detail under Materials and methods. Each experiment was performed in triplicate and the results are mean  $\pm$  SD. Values in the column are significantly different ( $P < 0.05$ )

<sup>a</sup>Total phenolics are expressed as gallic acid equivalent (GAE)

Incubation of the supercoiled form of  $\Phi$ X174 RF1 DNA with both H<sub>2</sub>O<sub>2</sub> and UV resulted in significant conversion of the supercoiled DNA to open circular form (lane 4). The prevention of DNA strand scission was detected when the DNA was incubated with 50–100  $\mu$ g/ml of heartwood extract (lanes 5 and 6). Strikingly, nearly complete protection by heartwood extract on the DNA scission was obtained at a dose of 500  $\mu$ g/ml (lane 7).

#### Total phenolics in the *Calocedrus formosana* extracts

The content of total phenolics in the extracts of *C. formosana* was determined by using the Folin–Ciocalteu assay and was expressed as gallic acid equivalents (GAE). According to the results shown in Table 1, the total phenolic contents of the heartwood, bark, and leaf extracts were 159.5  $\pm$  1.9, 115.3  $\pm$  2.3, and 53.5  $\pm$  1.5 mg/g, respectively. In other words, approximately 1.4-fold and three-fold higher amounts of phenolics existed in the heartwood extract relative to that of the bark and leaf extracts, respectively.

## Discussion

Various reactive oxygen species (ROS) encompass a spectrum of diverse chemical species including superoxide anions, hydrogen peroxide, hydroxyl radicals, nitric oxide, peroxynitrite, and others. These oxidants can play a variety

of roles in both animals and plants in vivo. For instance, some of the ROS are involved in cellular signaling, cell growth regulation, specific cellular physiology, and energy production;<sup>18</sup> however, the oxidation of lipids, DNA, protein, and carbohydrate by toxic ROS can also often cause DNA mutation, damage target cells or tissues, and result in cellular senescence and death. Recently, knowledge and application of potential antioxidant activities in reducing oxidative stresses in vivo have prompted many investigators to search for potent natural antioxidants from various plant sources.<sup>19–22</sup> *Calocedrus formosana*, the target plant of this study, is an indigenous plant species of Taiwan, and little or no systematic study has been performed to address the specific bioactivities of this plant related to its potential pharmacological or nutritional usage. Thus, in this study, we characterized not only the antioxidant activities but also the protective effect on the DNA scission for the plant extracts of *C. formosana*.

Based on the results of DPPH-radical assay, the ethanolic extracts of both heartwood and bark of *C. formosana* exhibited a significant inhibitory activity against DPPH radical, whereas the leaf extract was found to have much less effect on free radical inhibition, as shown in Fig. 1. The superoxide radical is known to be produced in vivo and can result in the formation of H<sub>2</sub>O<sub>2</sub> via dismutation reaction. Moreover, the conversion of superoxide and H<sub>2</sub>O<sub>2</sub> into more reactive species, e.g., the hydroxyl radical, has been thought to be one of the unfavorable effects caused by superoxide radicals.<sup>18</sup> Hence, in this report we have also evaluated the effect of *C. formosana* extracts on the inhibition of superoxide radicals. The data in Fig. 2 indicate that among the tested samples, the heartwood extract had the highest inhibitory activity against superoxide radicals induced by hypoxanthine-xanthin oxidase. Strikingly, the IC<sub>50</sub> value against the superoxide radical in heartwood extract is 2.3 times lower than that of (+)-catechin, a well-known antioxidant. The leaf extract showed the lowest activity on superoxide radical inhibition.

Previous reports have demonstrated that the phenolic content in plants can be correlated to their antioxidant activities.<sup>17,23,24</sup> In this study, the total phenolic content in the ethanol extracts of *C. formosana* was determined spectrometrically according to the Folin–Ciocalteu method and calculated as gallic acid equivalents (GAE), with the results as shown in Table 1. The total phenolic content of heartwood extract (159.5  $\pm$  1.9 mg/g) was higher than that of the bark extract (115.3  $\pm$  2.3 mg/g) and leaf extract (53.5  $\pm$  1.5 mg/g). These results suggest that the antioxidant activities of the heartwood, bark, and leaf extracts of *C. formosana* correlate well with their phenolic contents. In other words, the highest content of phenolics in heartwood extract has the most potent effect on antioxidant activity, relative to both bark and leaf extracts, as judged by the free radical and superoxide radical scavenging effects.

Hydroxyl radicals are considered to be the most active ROS, which can physically attack DNA to cause strand scission. Because the heartwood extract of *C. formosana* possessed the strongest antioxidant activity, we further characterized its protective activity on DNA strand scission.

As the data in Fig. 3 show, heartwood extract, at a dose of 1 mg/ml, virtually completely inhibited the  $\Phi$ X174 RF1 DNA strand cleavage under oxidative stress of hydroxyl radical induced by UV photolysis of  $H_2O_2$ . Keum et al.<sup>16</sup> have demonstrated that the methanolic extract of heat-processed ginseng can almost completely protect DNA from strand scission at a dose of 30 mg/ml. In contrast, a dosage of 30-fold less was required for the heartwood extract of *C. formosana* to inhibit DNA damage, compared to the methanolic extract of heat-processed ginseng. This methanolic extract from ginseng has also been found to have an inhibitory effect on mouse skin tumors, which was proposed to be associated with its antioxidant activity.<sup>16</sup>

It is now recognized that ROS may play major roles in tumor promotion and progression, chronic diseases, and aging.<sup>5</sup> The use of natural antioxidants as a potential preventive for free-radical mediated diseases has become a very important issue for improving the quality of life. This study demonstrates the significant antioxidant activity of plant extracts from *C. formosana*, especially the heartwood extract. This may indicate that the plant extracts or the derived phytochemicals from *C. formosana* have a great potential cancer chemopreventive effects or efficacies on other radical-mediated diseases. More systematic studies on phytochemistry as well as the in vitro and in vivo bioassays of *C. formosana* are currently in progress.

**Acknowledgments** This study was supported by a grant (NSC-91-2313-B-002-380) from the National Science Council of the Republic of China. The authors thank the National Science Council for this support and Mr. Tso-Yuan Hsieh (The Experimental Forest of National Taiwan University) for providing *Calocedrus formosana* materials.

## References

- Finkel T, Holbrook NJ (2000) Oxidants, oxidative stress and the biology of aging. *Nature* 408:239–247
- Pietta PG (2000) Flavonoids as antioxidants. *J Nat Prod* 63:1035–1042
- Cerutti PA (1994) Oxy-radicals and cancer. *Lancet* 344:862–863
- Frei B (1995) Cardiovascular disease and nutrient antioxidants: role of low-density lipoprotein oxidation. *Crit Rev Food Sci Nutr* 35:83–98
- Shireiqi I, Reddy P, Brenner DE (2000) Chemoprevention: general perspective. *Crit Rev Oncol/Hematol* 33:157–167
- Lin YT, Lo TB, Lin TH (1956) Study of the extractives constituents from the wood of *Libocedrus formosana* Florin. II. Interconversion between isoshonanonic acid and thujic acid. *J Chin Chem Soc* 3:36–40
- Lo TB, Lin YT (1956) Study of the extractives constituents from the wood of *Libocedrus formosana* Florin. I. *J Chin Chem Soc* 3:30–35
- Cheng YS, Lo TB, Chang LH, Lin YT (1961) Study of the extractives constituents from the wood of *Libocedrus formosana* Florin. IV. Partial hydrogenation of thujic acid. *J Chin Chem Soc* 8:103–108
- Fang JM, Jan ST, Cheng YS (1985) (+)-Calocedrin, a lignan dihydroanhydride from *Calocedrus formosana*. *Phytochemistry* 24:1863–1864
- Fang JM, Jan ST, Cheng YS (1987) Terpenoids from *Calocedrus formosana*. *Phytochemistry* 26:853–854
- Chang ST, Wang SY, Wu CL (2000) Identification of exudates from the bark of *Calocedrus formosana* (Florin) Florin (in Chinese). *For Prod Ind* 9:59–66
- Wang SY, Wu JH, Shyur LF, Kuo YH, Chang ST (2002) Antioxidant activity of abietane-type diterpenes from heartwood of *Taiwania cryptomerioides* Hayata. *Holzforchung* 56:487–492
- Pietta P, Simonetti P, Mauri P (1998) Antioxidant activity of selected medicinal plants. *J Agric Food Chem* 46:4487–4490
- Gyamfi MA, Yonamine M, Aniya Y (1999) Free-radical scavenging action of medicinal herbs from Ghana *Thonningia sanguinea* on experimentally-induced liver injuries. *Gen Pharmacol* 32:661–667
- Kirby AJ, Schmidt RJ (1997) The antioxidant activity of Chinese herbs for eczema and of placebo herbs-1. *J Ethnopharmacol* 56:103–108
- Keum YS, Park KK, Lee JM, Chun KS, Park JH, Lee SK, Kwon H, Surh YJ (2000) Antioxidant and anti-tumor promoting activities of the methanol extract of heat-processed ginseng. *Cancer Lett* 150:41–48
- Kujala TS, Loponen JM, Klika KD, Pihlaja K (2000) Phenolics and betacyanins in red beetroot (*Beta vulgaris*) root: distribution and effect of cold storage on the content of total phenolics and three individual compounds. *J Agric Food Chem* 48:5338–5342
- Halliwell B (1999) Food-derived antioxidants. Evaluating their importance in food and in vivo. *Food Sci Agric Chem* 1:67–109
- Zi X, Mukhtar H, Agarwal R (1997) Novel cancer chemopreventive effects of a flavonoid antioxidant silymarin: inhibition of mRNA expression of an endogenous tumor promoter TNF $\alpha$ . *Biochem Biophys Res Commun* 239:334–339
- Ohsugi M, Fan W, Hase K, Xiong Q, Tezuka Y, Komatsu K, Namba T, Saitoh T, Tazawa K, Kadota S (1999) Active-oxygen scavenging activity of traditional nourishing-herb medicinal and active constituents of *Rhodiola sacra*. *J Ethnopharmacol* 67:111–119
- Liu F, Ng TB (2000) Antioxidative and free radical scavenging activities of selected medicinal herbs. *Life Sci* 66:725–735
- Hu C, Kitts DD (2000) Studies on the antioxidant activity of *Echinacea* root extract. *J Agric Food Chem* 48:1466–1472
- Sato M, Ramarathnam N, Suzuki Y, Ohkubo T, Takeuchi M, Ochi H (1996) Varietal differences in the phenolic content and superoxide radical scavenging potential of wines from different sources. *J Agric Food Chem* 44:37–41
- Lavelli V, Peri C, Rizzolo A (2000) Antioxidant activity of tomato products as studied by model reactions using xanthine oxidase, myeloperoxidase, and copper-induced lipid peroxidation. *J Agric Food Chem* 48:1442–1448

The publication of this article was made possible by an Emachu Research Fund. The authors are grateful for the fund.