Antidyslipidemic Activity of Hot-water Extracts from Leaves of Cinnamomum osmophloeum Kaneh

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The antidyslipidemic activity of hot-water extracts of Cinnamomum osmophloeum leaves (COE) were evaluated on hamsters fed a high-fat diet. Oral administration of COE to hyperlipidemic hamsters reduced the total cholesterol (TC), triglyceride (TG) and low-density lipoprotein (LDL-C) levels. Compared with hyperlipidemic hamsters, the plasma TC and TG levels of hamsters fed with COE at a dosage of 100 mg/kg body weight for 5 and 10 weeks were significantly reduced to 12.63% and 34.25%, and 33.88% and 36.88%, respectively. Plasma LDL-C was also reduced to 27.77% after 10 weeks feeding with the same regimen. Standard diagnostic tests indicated that the extracts did not cause damage to hamster liver or kidneys. Based on these results, it is concluded that COE possesses antidyslipidemic activity. The composition of COE was characterized. Two main compounds, kaempferol 3-O-β-d-apiofuranosyl-(1→2)-α-L-arabinofuranosyl-7-O-α-L-rhamnopyranoside (1) and kaempferitin (2) were identified in the hot-water extracts. Their contents were 7.56% and 9.95%, respectively. Copyright © 2011 John Wiley & Sons, Ltd.

Keywords: Cinnamomum osmophloeum Kaneh; hot-water extract; total cholesterol; triglyceride; flavonol glycosides; antidyslipidemic activity.

INTRODUCTION

The genus Cinnamomum comprises about 250 species that are indigenous to Asia and Australia (Cheng et al., 2006). The native name for Cinnamomum osmophloeum Kaneh (Lauraceae) is ‘indigenous cinnamon tree’. It is an endemic hardwood grown in Taiwan at elevations of 400–1500 m (Chao et al., 2005). One distinguishing characteristic of C. osmophloeum is that the composition of the essential oil of the leaves is similar to that of C. cassia. Several previous studies have demonstrated that the essential oil of C. osmophloeum leaves possesses a broad range of bioactivity against different kinds of organisms including bacteria (Cheng et al., 2004), termites (Chang et al., 2001), mites (Chen et al., 2002), mildew (Chen and Chang, 2002) and fungi (Wang et al., 2005). Recently, Wang and coworkers (Wang et al., 2008) found that the essential oil and its dominant compound, cinnamaldehyde, possessed strong xanthine oxidase inhibitory activity and had an antihyperuricemia effect in mice. Furthermore, Sheng and his co-workers approved that cinnamaldehyde possessed the activation activity for peroxisome proliferator-activated receptors γ and α (PPARγ/α), and could improve insulin resistance, and to reduce fasted glucose, FFA, LDL-C, and AST levels in high-caloric diet-induced obesity (DIO) and db/db mice (Sheng et al., 2008). They also demonstrated the hot-water extract prepared from cinnamon bark could increase the expression of PPARγ/α and their target genes such as LPL, CD36, GLUT4 and ACO in 3T3-L1 adipocyte in vitro (Sheng et al., 2008).

In addition to essential oil, the leaves of C. osmophloeum contain abundant flavonol glycosides (Fang et al., 2005). Epidemiological studies indicate that flavonoids and flavonol glycosides have various biological activities, such as decreasing the risk of cardiovascular diseases (Woodman and Chan, 2004), cancer chemoprevention activity (Rao et al., 2009) and cholesterol-lowering effects (Kurowska and Manthey, 2004). In addition, there is considerable evidence suggesting a relationship between dyslipidemia and the manifest characteristics of metabolic syndrome X. Dyslipidemia has been well documented and is characterized by an ‘atherogenic lipoprotein phenotype’ or ‘lipid triad’. Low-density lipoprotein (LDL) is one of the five major groups of lipoproteins, which in order of size, largest to smallest, are chylomicrons, VLDL, IDL, LDL and HDL, that enable lipids such as cholesterol and triglycerides to be transported within the water-based bloodstream. Medically, estimates of cholesterol content carried by LDL particles are used as an index to predict the blood cholesterol condition. Besides, higher levels of LDL particles bring about medical problems such as cardiovascular disease, they are often called the bad cholesterol particles (Colpo,
Plant extract preparation. Air-dried and powdered leaves (950 g) of *C. osmophloeum* were extracted by boiling for 1 h with 15 L hot distilled water. After filtration with cheesecloth, the filtrates were collected and concentrated by lyophilization, yielding 150 g of COE (yield = 16%, w/dry weight of leaves).

Plant materials and chemicals. The leaves of *C. osmophloeum* were collected in May 2009 at Hui-Sun Experimental Forest Station, which is located in the middle of Taiwan. The plants were authenticated by Dr Yen-Hsueh Tseng (Professor of Department of Forestry, National Chung-Hsing University). The voucher specimen [Tseng4518] was deposited in the herbarium of the same university. All chemicals and solvents used for the HPLC grade. Deionized water was prepared using a Milli-Q water purification system (Millipore, MA). All the solutions were filtered through 0.45 μm membranes (Schleicher & Schuell, Germany) and degassed by an ultrasonic bath before use. Gemfibrozil with a purity ≥99%, purchased from Sigma-Aldrich (StLouis, MO, USA), was used as the reference cholesterol lowering drug.

Animals and experiment protocol. Animal experiments were designed in accordance with the *Guidebook for the Care and Use of Laboratory Animals* of The Chinese Taipei Society of Laboratory Animal Science and approved by the same society. Forty male Golden Syrian hamsters 7 weeks old, weighing 100–120 g were housed in individual plastic cages and subjected to a 12 h light/dark cycle, relative humidity of 60% and a temperature at 25 °C. Experimental diets were provided a standard laboratory diet (LabDiet® 5001 Rodent diet, Purina Mills LLC, St Louis, MO, USA). The animals were given free access to regular rodent chow and water. Food intake was recorded daily and the animals were weighed weekly. The dosages of leaf extracts and gemfibrozil were adjusted weekly according to the average body weight of each hamster. Hamsters were fed a high-cholesterol diet and orally administered *C. osmophloeum* leaf extracts at dosages of 500 mg/kg body weight (bw) per day (HChol-CO500), 100 mg/kg bw per day (HChol-CO100). The positive control group (HChol-gem20) was fed a high-cholesterol diet and orally administered 20 mg/kg bw per day of gemfibrozil. The hamsters were fed these experimental diets for 5 and 10 weeks. At the end of the treatment period, the animals were anesthetized and killed by inhalation of 2% isoflurane, and blood samples were collected from the orbital sinus and cardiac puncture. Plasma samples were then prepared. All plasma samples were used for analysis of total cholesterol (TC), triglyceride (TG), HDL-cholesterol (HDLC), LDL-cholesterol (LDLC), creatinine and blood urea nitrogen (BUN). All analyses were carried out in triplicate according to the spectrophotometry of Chiron Diagnostics Corporation (Oberlin, OH, USA) using the Express Plus Automatic Clinical Chemistry Analyzer (Chiron).

Compound isolation and index compound quantification. The COE was separated by analytical high-performance liquid chromatography (HPLC) using an Agilent1100 (Agilent, Germany) system with a 100 μL fixed loop, and a UV detector. A Luna C18 column (4.60 mm × 250 mm, 5 μm particle size, Phenomenex, Torrance, CA) with a three solvent system, 1% phosphoric acid with deionized water (solvent A), methanol (solvent B) and acetonitrile (solvent C). The linear gradient program was as follows: 82% A, 3% B and 15% C in the first 0 min, held for 10 min, changed to 77% A, 3% B and 20% C over 40 min, then changed to 100% B at 30 min, held for 5 min. The mobile phase flow rate was 0.4 mL/min, and the detector was monitored at 254 nm. All the chromatographic operations were carried out at ambient temperature (25 °C). Two major compounds were obtained from the hot-water extracts at retention times of 36.0 min (1) and 37.9 min (2). The structures of 1 and 2 were then elucidated using spectroscopic analysis. UV and IR data were acquired on a Bio-Tek μQuant MX200 Microplate Spectrophotometer and a PerkinElmer Spectrum 100 FT-IR spectrometer, respectively. 1H-NMR and 13C-NMR spectra were obtained on a Varian Unity Inova-600 MHz spectrometer using DMSO-d6 as solvent. The HREIMS data were performed with a Thermo/Finnigan Quest MAT 95XL mass spectrometer. For calibration curves, appropriate volumes of the standard stock solutions (2 mg/mL) were diluted with deionized water, and five concentration levels (100, 250, 500, 1000 and 2000 μg/mL) were analysed. For quantification, the peak areas were correlated with the concentrations according to the calibration curve. All data are expressed as mean ± standard deviation of triplicate independent experiments (n = 3).

Statistical analysis. The main treatment effects were analysed by one-way analysis of variance (ANOVA) followed by Duncan’s multiple range test using a Statistical Analysis System software (SAS Institute, Cary, NC). The significance level was set at p < 0.05. Data were expressed as mean ± SD.
RESULTS

Effects of C. osmophloeum on body weights of hamsters

This study investigated the effects of oral administration of hot water extract of C. osmophloeum leaves (COE) on a hyperlipidemic hamster model. During the experiment period, the body weights and daily food intake of the hamsters increased normally and did not show any difference among the various groups (Table 1). All the experimental animals had normal physical appearance and health throughout the experimental period. At weeks 5 and 10 after the start of the treatment regimen the animals were anesthetized and blood samples were collected and examined for estimation of the hypolipidemic effects of COE.

Regulation effects of C. osmophloeum on TC and TG levels

Table 2 shows the changes in plasma TC and TG levels. At the time for starting treatment with water extracts and gemfibrozil (week 0), the control group had a TC of 83.6 ± 8.2 mg/dL and the HChol group had extremely high TC levels of 219.8 ± 24.2 mg/dL. The high TC levels of the HChol group hamsters in comparison with the control group indicated that the HChol group was suitable for use as an animal model for antihyperlipidemic assay. The results revealed that the plasma TC and TG of hamsters were lowered by daily feeding with hot water extracts of C. osmophloeum at dosages of 100 mg/kg bw per day (HChol-CO100) and 500 mg/kg bw per day (HChol-CO500) for 5 and 10 weeks. The HChol-CO100 group had plasma TC and TG concentrations of 197.8 ± 47.2 and 126.1 ± 34.1 mg/dL after 5 weeks of treatment, which were 12.6% and 30.9%, respectively, less than those of the HChol group (p < 0.05). At week 10, the plasma TC and TG levels of HChol-CO100 were 153.2 ± 25.5 and 137.2 ± 78.6 mg/dL, which was less than 34.2% and 33.9%, respectively, of the levels in the HChol group. In the higher dosage group (HChol-CO500) more pronounced effects were observed. At week 10, the plasma TC and TG levels of HChol-CO500 were decreased to 40.3% (139.2 ± 17.2 ng/dL and 120.8 ± 49.3 ng/dL). After 5 and 10 weeks, gemfibrozil, a proven cholesterol reducer used clinically, significantly reduced the plasma TC levels of the hyperlipidemic hamsters (p < 0.05) (Khan et al., 2008).

Changes of LDL-C/HDL-C ratio in the serum by treating with C. osmophloeum extracts

Effects of COE on serum HDL-C and LDL-C levels of hamsters are shown in Fig. 1A. The LDL-C levels of hamsters were significantly higher in the HChol group (126.9 ± 40.1 mg/dL) than in the control (53.4 ± 8.1 mg/dL). The LDL-C levels of the HChol group hamsters were decreased to 12.6% and 50.9%, respectively, less than those of the control group (p < 0.05). At week 10, the LDL-C levels of HChol-CO100 were 34.2% and 33.9%, respectively, of the levels in the HChol group.

Table 1. Hamster body weight and daily food intake gain

<table>
<thead>
<tr>
<th>Group</th>
<th>Body weight (g)</th>
<th>Daily feed intake (g/day)</th>
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</thead>
<tbody>
<tr>
<td></td>
<td>Week 0</td>
<td>Week 5</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>100.4 ± 3.3^a</td>
<td>121.2 ± 4.2^a</td>
</tr>
<tr>
<td>HChol</td>
<td>99.3 ± 4.0^a</td>
<td>114.8 ± 3.7^a</td>
</tr>
<tr>
<td>HChol-Gem20</td>
<td>101.0 ± 3.4^a</td>
<td>116.7 ± 5.4^a</td>
</tr>
<tr>
<td>HChol-CO100</td>
<td>101.0 ± 2.0^a</td>
<td>117.1 ± 5.9^a</td>
</tr>
<tr>
<td>HChol-CO500</td>
<td>100.5 ± 1.9^a</td>
<td>114.8 ± 6.8^a</td>
</tr>
</tbody>
</table>

Control, normal diet (0% cholesterol); HChol, high-cholesterol diet; HChol-Gem20, gemfibrozil 20 mg/kg body weight (bw) per day and high-cholesterol diet; HChol-CO100, COE 100 mg/kg bw per day and high-cholesterol diet; HChol-CO500, COE 500 mg/kg bw per day and high-cholesterol diet. Data are presented as mean ± SD (n = 8). Mean values within each column with different labels (a,b,c) are significantly different (p < 0.05).

Table 2. Effects of COE on hamster plasma TC and TG levels

<table>
<thead>
<tr>
<th>Group</th>
<th>Cholesterol (mg/dL)</th>
<th>Triglyceride (mg/dL)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Week 0</td>
<td>Week 5</td>
</tr>
<tr>
<td></td>
<td>(mg/dL)</td>
<td>(mg/dL)</td>
</tr>
<tr>
<td>Control</td>
<td>85.6 ± 8.2^a</td>
<td>85.0 ± 19.6^a</td>
</tr>
<tr>
<td>HChol</td>
<td>219.8 ± 24.2^b</td>
<td>232.9 ± 92.9^c</td>
</tr>
<tr>
<td>HChol-Gem20</td>
<td>235.7 ± 17.2^b</td>
<td>148.1 ± 41.7^ab</td>
</tr>
<tr>
<td>HChol-CO100</td>
<td>243.5 ± 36.3^b</td>
<td>147.0 ± 43.9^ab</td>
</tr>
<tr>
<td>HChol-CO500</td>
<td>237.0 ± 48.6^b</td>
<td>208.8 ± 79.7^ab</td>
</tr>
</tbody>
</table>

Control, normal diet (0% cholesterol); HChol, high-cholesterol diet; HChol-Gem20, gemfibrozil 20 mg/kg bw per day and high-cholesterol diet; HChol-CO100, COE 100 mg/kg bw per day and high-cholesterol diet; HChol-CO500, COE extracts 500 mg/kg bw per day and high-cholesterol diet. Data are presented as mean ± SD (n = 8). Mean values within each column with different labels (a,b,c) are significantly different (p < 0.05).
Effects of *C. osmophloeum* extracts on serum GOT, GPT, BUN and creatinine levels

It was demonstrated that COE was able to decrease plasma TC and TG levels effectively, but whether it had other adverse effects on hamsters was still unclear. Therefore glutamate oxaloacetate transaminase (GOT) and glutamic pyruvic transaminase (GPT) levels were measured. GOT and GPT are released into the blood from liver cells when the cells are damaged and are regularly used to diagnose liver and heart disease. Plasma GOT and GPT levels in hyperlipidemic hamsters did not increase when they were fed COE (Table 3). However, the HChol group had higher plasma GOT and GPT levels than the control group after 10 weeks. In addition, the HChol group revealed a higher plasma blood urea nitrogen (BUN) level (32.0 ± 11.2 mg/dL) than the control group (25.1 ± 2.9 mg/dL). But there was no significant difference between the COE treated groups and control. Similar results were also found for creatinine levels in plasma (Table 3).

Characterization of composition of hot-water extracts of *C. osmophloeum* leaves

Finally, the composition of COE was analysed by HPLC. Figure 2 shows the HPLC chromatogram of COE. Two main compounds (compounds 1 and 2) were isolated from the hot-water extract and characterized. According to the MS and NMR analyses, compounds 1 and 2 were identified as kaempferol 3-O-β-D-apiofuranosyl-(1→2)-α-L-arabinofuranosyl-7-O-α-L-rhamnopyranoside (1) and kaempferitin (2); spectral data were consistent with the literature (Rao et al., 2005). The standard calibration curves (peak area vs concentration) of compound 1 and compound 2, were in the range 100–2000 μg/mL. The linear regression curves were $y = 29.25x + 2298.8$ ($R^2 = 0.9941$) for compound 1 and $y = 23.83x + 2833.8$ ($R^2 = 0.9852$) for compound 2 and revealed a good linearity. According to HPLC analysis, the contents of compounds 1 and 2 in COE were 7.56% and 9.95%, respectively.

DISCUSSION

The antidysslipidemic activity of hot-water extracts from the leaves of *C. osmophloeum* was evaluated in this study. According to the results obtained, *C. osmophloeum* extracts possessed significant reducing effects on TC, TG and LDL-C without changing the LDL-C/HDL-C ratio in hamster plasma. If the LDL-C/HDL-C ratio was lowered, then the content of HDL-C had a much higher percentage in TC levels, while on the contrary, the atherosclerotic risk factor LDL-C was lowered. Even administered at a low dosage (100 mg/kg bw per day) of hot-water extract to the hamsters, it expressed a remarkable effect on cholesterol-lowering activity.
Table 3. Plasma level change of HDL-C, LDL-C, GOT, GPT, BUN and creatinine of hamsters fed with COE for 10 weeks

<table>
<thead>
<tr>
<th>Group</th>
<th>GOT (U/dL)</th>
<th>GPT (U/dL)</th>
<th>BUN (mg/dL)</th>
<th>Creatinine (mg/dL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>43.0 ± 7.5*</td>
<td>65.9 ± 11.8*</td>
<td>25.1 ± 2.9ab</td>
<td>0.3 ± 0.1b</td>
</tr>
<tr>
<td>HChol</td>
<td>99.6 ± 34.3b</td>
<td>127.6 ± 19.0b</td>
<td>32.0 ± 11.2b</td>
<td>0.3 ± 0.0b</td>
</tr>
<tr>
<td>HChol-Gem20</td>
<td>49.6 ± 5.6a</td>
<td>70.2 ± 13.3a</td>
<td>19.3 ± 3.6a</td>
<td>0.2 ± 0.0a</td>
</tr>
<tr>
<td>HChol-CO100</td>
<td>62.9 ± 33.7ab</td>
<td>79.5 ± 30.8a</td>
<td>22.0 ± 2.3a</td>
<td>0.2 ± 0.1ab</td>
</tr>
<tr>
<td>HChol-CO500</td>
<td>53.1 ± 11.7*</td>
<td>71.5 ± 18.8*</td>
<td>18.8 ± 2.9*</td>
<td>0.2 ± 0.0*</td>
</tr>
</tbody>
</table>

Control, normal diet (0% cholesterol); HChol, high-cholesterol diet; HChol-Gem20, gemfibrozil 20 mg/kg bw per day and high-cholesterol diet; HChol-CO100, COE 100 mg/kg bw per day and high-cholesterol diet; HChol-CO500, COE 500 mg/kg bw per day and high-cholesterol diet. Data are presented as mean ± SD (n = 8). Mean values within each column with different labels (a, b, c) are significantly different (p < 0.05).

Moreover, the lipid concentration in plasma was reduced in a dose dependent manner by treatment with hot-water extracts of *C. osmophloeum* leaves. More cholesterol supplementation will lead to a burden on the liver metabolites and simultaneously increase the risk for developing hepatic fibrosis (Aguilera et al., 2005; Jeong et al., 2005; Papadia et al., 2004). According to the results observed in this study, the LDL-C levels in plasma of hamsters treated with extract were lower than those of the high cholesterol uptake only group (HChol). However, more investigations are needed to ascertain this effect. From the perspective of food science, a dosage of 100 mg/kg bw in the hamster corresponds to supplementing the diet of a human with a body weight of 70 kg and a height of 170 cm with COE at 854 mg daily (Reagan-Shaw et al., 2008). Even administered at this low dosage as a hot-water extract, the leaves of *C. osmophloeum* expressed a remarkable cholesterol-lowering activity in the hamster. It is concluded that the COE might be a safe food supplement for treating dyslipidemia. Further investigations into bioavailability, clinical safety and so on should be conducted in the future.

Conflict of Interest

The authors have declared that there is no conflict of interest.

REFERENCES


Gray RS, Robbins DC, Wang W et al. 1997. Relation of LDL size to the insulin resistance syndrome and coronary heart disease in


