

Genetic relationships of *Rhododendron* (*Ericaceae*) in Taiwan based on the sequence of the internal transcribed spacer of ribosomal DNA

By C. C. TSAI^{1,2}, S. C. HUANG³, C. H. CHEN⁴, Y. H. TSENG⁴, P. L. HUANG¹, S. H. TSAI³ and C. H. CHOU^{2,5*}

¹Kaohsiung District Agricultural Improvement Station, Pingtung 900, Taiwan

²National Sun Yat-sen University, Kaohsiung 804, Taiwan

³Taichung District Agricultural Improvement Station, Changhwa 515, Taiwan

⁴Taiwan Endemic Species Research Institute, Nantou 552, Taiwan

⁵National Pingtung University of Science and Technology, Pingtung 912, Taiwan

(e-mail: choumasa@mail.npust.edu.tw)

(Accepted 25 November 2002)

SUMMARY

The genetic relationships of 20 *Rhododendron* species in Taiwan was determined based on the sequence of the internal transcribed spacer (ITS) region of ribosomal DNA. Sequences of the complete ITS region including ITS1, 5.8S rDNA, and ITS2, were obtained by direct sequencing of polymerase chain reaction (PCR)-amplified fragments. *Gaultheria itoana* was used as an outgroup. Aligned sequences of ITS1 and ITS2 from the 21 taxa resulted in 493 characters. According to the dendrogram, six main clusters were classified among the 20 species of the genus *Rhododendron* in Taiwan. *Rhododendron oldhamii*, *R. nakaharai*, *R. taiwanalpinum*, *R. simsii*, *R. lasiostylum*, *R. rubropilosum*, *R. breviperulatum*, *R. kanehirai*, and *R. noriakianum* were grouped with *R. longiperulatum* in cluster I. *Rhododendron lamprophyllum* was grouped with *R. ovatum* in cluster II. *Rhododendron pseudochrysanthum*, *R. morri*, *R. hyperythrum*, and *R. rubropunctatum* were grouped with *R. formosanum* in cluster III. In addition, *R. mariesii*, *R. ellipticum* and *R. kawakamii* formed three independent clusters. In this study, the findings based on ITS sequences are in agreement with the systematics of *Rhododendron*.

Azaleas (*Rhododendron* spp.) are horticulturally important ornamental plants and have been extensively hybridized (Kron *et al.*, 1993). Approximately, 850 native azaleas species have been found in the world (Sleumer, 1966). Over 1000 horticultural hybrids had been bred by artificial hybridization to 1976 (Bean, 1976). Variability within species and an absence of distinguishing morphological characteristics between species have caused difficulties in defining taxa of azalea plants (Rayburn *et al.*, 1993). Therefore, only 500 to 600 species have been identified within the genus *Rhododendron* based on different standpoints of classification (Willis, 1985).

The genus *Rhododendron* has been subdivided into eight subgenera (Kron and Judd, 1990). Four subgenera, including 20 species, were identified within the genus *Rhododendron* in Taiwan (Li, 1978; He *et al.*, 1999; Yang *et al.*, 1999). A taxonomic revision of this genus was conducted based on morphology, habitat, and flowering season in Taiwan. Others have suggested that only 14 species and one doubtful species occur within the genus *Rhododendron* in Taiwan (Lu and Yang, 1989).

Plant taxa of the *Ericaceae* show a great deal of morphological variability and have been thoroughly studied based on characteristics of morphology, embryology, anatomy, and chemistry (Kron *et al.*, 1999). Morphological characters have traditionally been used to distinguish species. However, most morphological characters are easily influenced by environmental factors (Iqbal *et al.*, 1995). Recently, molecular data have been

introduced to assess several higher-level phylogenetic analyses within the *Ericaceae* or Ericales. For example, *rbcL* gene sequences of chloroplast DNA (cpDNA) were useful for elucidating the systematics of the *Ericaceae*, *Empetraceae*, and *Epacridaceae* (Kron and Chase, 1993); 18S rRNA gene sequences were useful for determining the phylogenetic relationships of the *Empetraceae*, *Epacridaceae*, *Ericaceae*, *Monotropaceae*, and *Pyrolaceae* (Kron, 1996); and *matK* gene sequences were used for determining the phylogenetic relationships of the *Lyonia* group of the *Andromedeae* (*Ericaceae*) (Kron and Judd, 1997), as well as within the *Andromedeae* (*Ericaceae*) (Kron *et al.*, 1999).

Because ITS regions of rDNA show great divergence, it has been suggested that ITS regions would be quite useful for comparing among closely related organisms or for the study of microevolutionary processes among or even within populations (Baldwin, 1993; Ritland and Straus, 1993). Sequence comparison of the ITS region of rDNA has been used to reconstruct the phylogeny of numerous plant taxa, such as *Calycadenia* (*Asteraceae*) (Baldwin, 1993), *Paeonia* (*Ranunculaceae*) (Sang *et al.*, 1995), and *Sorghum* (*Poaceae*) (Sun *et al.*, 1994). The ITS sequence also has been used to determine the genetic relationship of the section *Pentanthera* (*Ericaceae*) (Scheiber *et al.*, 2000).

In this study, PCR-amplified products of direct DNA sequencing were used to determine the ITS sequences of rDNA of the 20 *Rhododendron* species in Taiwan. The genetic relationships among these 20 *Rhododendron* species was thus established based on their ITS sequences.

*Author for correspondence.

TABLE I
Twenty taxa of *Rhododendron* spp. used in this study and their systematic classification, ITS length, and accession number

Species and classification	Source ^a	ITS length	Accession no. in GenBank
Subgenus <i>Hymenanthes</i>			
Section <i>Vireya</i>			
<i>R. formosanum</i>	A	642	AF285854
<i>R. hyperythrum</i>	A	642	AF432426
<i>R. morri</i>	A	642	AF432422
<i>R. pseudochrysanthum</i>	A	642	AF432424
<i>R. rubropunctatum</i>	B	642	AF285842
Subgenus <i>Tsutsusi</i>			
Section <i>Tsutsusi</i>			
<i>R. breviperulatum</i>	A	643	AF432425
<i>R. kanehirai</i>	B	643	AF172290
<i>R. lasiostylum</i>	B	643	AF285845
<i>R. longiperulatum</i>	B	643	AF285847
<i>R. nakaharai</i>	B	643	AF285846
<i>R. noriakianum</i>	A	643	AF285856
<i>R. oldhamii</i>	B	643	AF285843
<i>R. rubropilsoum</i>	B	643	AF285849
<i>R. simsii</i>	B	643	AF285848
<i>R. taiwanalpinum</i>	A	643	AF432479
Section <i>Brachycalyx</i>			
<i>R. mariesii</i>	B	643	AF285844
Subgenus <i>Azaleastrum</i>			
Section <i>Azaleastrum</i>			
<i>R. lamprophyllum</i>	A	643	AF285855
<i>R. ovatum</i>	A	643	AF432421
Section <i>Choniastrum</i>			
<i>R. ellipticum</i>	B	643	AF285841
Subgenus <i>Rhododendron</i>			
Section <i>Vireya</i>			
<i>R. kawakamii</i>	A	648	AF432450

^a A = from the Taiwan Endemic Species Research Institute, B = from a transplant nursery.

MATERIALS AND METHODS

Total DNA extraction

Twenty species of *Rhododendron* in Taiwan plus a single species, *Gaultheria itoana* from another genus, as an outgroup, were used in this study (summarized in Table I). Total cellular DNA was extracted from fresh or silica-gel dried (Chase and Hillis, 1991) leaves, using the method of CTAB (cetyltrimethylammonium bromide) (Doyle and Doyle, 1987). Approximate DNA yields were determined using a spectrophotometer (Hitachi U-2001), and the DNA samples were then stored in a freezer at -20°C .

Primer design and PCR amplification

One primer was designed from the conserved regions of the 5' end of 18S rDNA of rice (Takaiwa *et al.*, 1984) and tomato (Kiss *et al.*, 1989a). Another primer was designed from their complementary conserved regions of the 3' end of 26S rDNA. In the present study, two sets of primers, i.e. IT1: 5'TCGTAACAAGGTTTCCGTAGGT3' and IT2: 5'GTAAGTTTCTTCTCCTCCGCT3' were designed to amplify the internal transcribed spacer (ITS) of rDNA in azalea plants (Figure 1). PCR was performed as follows: we used a 50 μl mixture containing 10 mM Tris-HCl (pH 9.0), 50 mM KCl, 2 mM MgCl_2 , 0.01% BSA, with four dNTPs (0.2 mM each), primers (0.5 μM each), 2.5 units of *Taq* DNA polymerase (Virogene), 10 ng genomic DNA, and a 50 μl volume of mineral oil. Amplification reactions were performed in a dry-block with two-step thermal cycles (Biometra). In the first step, the mixture was incubated at 94°C for 3 min, then it underwent 10 cycles of denaturation at 94°C for 45 s, annealing at 58°C for 45 s, and extension at 72°C for 1 min. The second step was carried out by the following process: 30 cycles of denaturation at 94°C for 45 s, annealing at 54°C for 45 s, extension at 72°C for 1 min, with a final extension

for 10 min at 72°C . These PCR products were detected by agarose gel electrophoresis (1.0%, w/v in TBE), and staining by $0.5 \mu\text{g/ml}^{-1}$ of ethidium bromide, and were finally photographed under UV light exposure.

DNA sequencing

PCR products of ITS sequences from the 20 *Rhododendron* species were recovered by glassmilk (BIO 101, California) and directly sequenced by the dideoxy chain-termination method using an ABI377 automated sequencer with a BigDyeTM Terminator Cycle Sequencing Ready Reaction Kit (PE Biosystems, California). Sequencing primers were the same as those used for PCR. Each sample was sequenced two or three times to confirm the sequences. These reactions were performed based on the recommendations of the manufacturers.

Data analysis

Genetic relationships were determined by using the program MEGA version 2.1 (Kumar *et al.*, 2001). The genetic distance matrix was calculated by the two-parameter method of Kimura (1980). These data were then used to construct a phylogenetic tree using the unweighted pair-group method analysis (UPGMA) (Rohlf *et al.*, 1982).

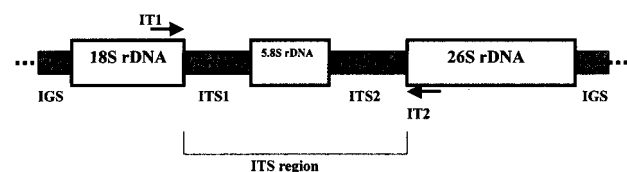


FIG. 1

The structure of ribosomal DNA. Positions of internal transcribed spacer (ITS) regions relative to 18S, and 26S rRNA genes and the intergenic spacer (IGS) are shown. The relative position of the primers used for PCR is indicated.

	280	290	300	310	320	330	340	350	360
<i>R. breviperulatum</i>	TCACCCCGTG	CCTCATCGGC	AGGTAAGTGC	GTGGGCGGAT	ATTGGCCCCC	CGTGCACATT	GGTGCTCGGC	CGGCCTAAAA	ATGACGGTCC
<i>R. ovatum</i>									
<i>R. formosanum</i>		A	G			T	C	T	
<i>R. hyperythrum</i>		A	G			T	C	T	
<i>R. kanehirai</i>									
<i>R. kawakamii</i>			G		A		T	C	T
<i>R. lambrophyllum</i>							T	C	
<i>R. lasiostylum</i>									
<i>R. longiperulatum</i>									
<i>R. mariesii</i>		A					T	C	T
<i>R. morri</i>		A	G				T	C	T
<i>R. nakaharai</i>									
<i>R. noriakianum</i>									
<i>R. oldhamii</i>									
<i>R. ellipticum</i>			G				T	C	T
<i>R. pseudochrysanthum</i>		A	G				T	C	T
<i>R. rubropilosum</i>									
<i>R. rubropunctatum</i>		A	G				T	C	T
<i>R. simsii</i>									
<i>R. taiwanalpinum</i>									

	370	380	390	400	410	420	430	440	450
<i>R. breviperulatum</i>	CCGATGACGG	ACATCACGGC	AAGTGGTGGT	TGCCAAACCG	TGCGTCATG	TCGTGCATGC	CA-TTCTTTG	TCGCGGGGCT	GGCTCATCGA
<i>R. ovatum</i>									
<i>R. formosanum</i>	T								
<i>R. hyperythrum</i>	T								
<i>R. kanehirai</i>									
<i>R. kawakamii</i>	A						A		G
<i>R. lambrophyllum</i>					G				
<i>R. lasiostylum</i>									
<i>R. longiperulatum</i>									
<i>R. mariesii</i>							T		
<i>R. morri</i>	T								
<i>R. nakaharai</i>									
<i>R. noriakianum</i>									
<i>R. oldhamii</i>									
<i>R. ellipticum</i>					GC			T	A
<i>R. pseudochrysanthum</i>	T								
<i>R. rubropilosum</i>									
<i>R. rubropunctatum</i>	T							TT	
<i>R. simsii</i>									
<i>R. taiwanalpinum</i>									

	460	470	480
<i>R. breviperulatum</i>	CCCTTAAGTA	CCATATAC--	-TGCGGTACC TCAACT
<i>R. ovatum</i>		CA	--T
<i>R. formosanum</i>		CA	--T
<i>R. hyperythrum</i>		CA	--T
<i>R. kanehirai</i>			
<i>R. kawakamii</i>		CA AA	C CT
<i>R. lambrophyllum</i>		CA	--T
<i>R. lasiostylum</i>			
<i>R. longiperulatum</i>			
<i>R. mariesii</i>		T CA	--T C
<i>R. morri</i>		CA	--T
<i>R. nakaharai</i>			
<i>R. noriakianum</i>			
<i>R. oldhamii</i>			
<i>R. ellipticum</i>		CA T	--T T
<i>R. pseudochrysanthum</i>		CA	--T
<i>R. rubropilosum</i>			
<i>R. rubropunctatum</i>		CA	--T
<i>R. simsii</i>			
<i>R. taiwanalpinum</i>			

FIG. 2

Sequence comparison of the ITS1 and ITS2 regions from 20 taxa of the genus *Rhododendron* and *G. itoana* in Taiwan. Dots (.) indicate the same nucleotides, and gaps (-) are introduced to maximize homology.

being more diverse, and the group used including members of four subgenera of *Rhododendron*.

Aligned sequences of ITS1 and ITS2 from the 21 taxa resulted in 493 characters. A distance matrix of genetic divergence values among the 21 taxa in this study is given in Table II. Genetic distances among the 20 *Rhododendron* species are in the range from 0 to 0.062 as measured by the two-parameter method described by Kimura (1980). In comparison with other species, extremely high divergence values in the range of from 0.190 to 0.204 were found among species of the genus *Rhododendron* and the outgroup *G. itoana*. The highest

genetic difference value of 0.204 was found between *G. itoana* and *R. ovatum* (Table II).

A dendrogram was obtained from ITS sequence comparisons. According to the dendrogram, the 20 *Rhododendron* species in Taiwan were grouped into six clusters. *Rhododendron olhamii*, *R. nakaharai*, *R. taiwanalpinum*, *R. simsii*, *R. lasiostylum*, *R. rubropilosum*, *R. breviperulatum*, *R. kanehirai*, and *R. noriakianum* were grouped with *R. longiperulatum* in cluster I. *Rhododendron lambrophyllum* was grouped with *R. ovatum* in cluster II. *Rhododendron pseudochrysanthum*, *R. morri*, *R. hyperythrum*, and *R. rubropunctatum* were grouped with *R. formosanum* in

TABLE II
Genetic distance of the ITS sequence among 20 species of the genus *Rhododendron* plus *Gaultheria itoana* according to the two-parameter method of Kimura

	<i>R. formosanum</i>	<i>R. hyperythrum</i>	<i>R. morri</i>	<i>R. pseudo-chrysanthum</i>	<i>R. rubropunctatum</i>	<i>R. breviperulatum</i>	<i>R. kanehirai</i>	<i>R. lasiosyrtum</i>	<i>R. longiperulatum</i>	<i>R. nakaharai</i>	<i>R. noriakianum</i>	<i>R. oldhamii</i>	<i>R. rubropilatum</i>	<i>R. sinii</i>	<i>R. taiwanalpinum</i>	<i>R. mariesii</i>	<i>R. lamprophyllum</i>	<i>R. ovatum</i>	<i>R. ellipticum</i>	<i>R. kawakamii</i>		
<i>R. formosomum</i>																						
<i>R. hyperythrum</i>	0.013																					
<i>R. morri</i>	0.013	0.000																				
<i>R. pseudo-chrysanthum</i>	0.013	0.000	0.000																			
<i>R. rubropunctatum</i>	0.017	0.004	0.004	0.004																		
<i>R. breviperulatum</i>	0.059	0.050	0.050	0.050	0.054																	
<i>R. kanehirai</i>	0.054	0.046	0.046	0.046	0.050	0.002																
<i>R. lasiosyrtum</i>	0.057	0.048	0.048	0.048	0.052	0.006	0.002															
<i>R. longiperulatum</i>	0.061	0.052	0.052	0.052	0.057	0.006	0.004	0.002														
<i>R. nakaharai</i>	0.057	0.048	0.048	0.048	0.052	0.002	0.000	0.004	0.004													
<i>R. noriakianum</i>	0.059	0.050	0.050	0.050	0.054	0.006	0.004	0.002	0.004	0.004												
<i>R. oldhamii</i>	0.059	0.050	0.050	0.050	0.054	0.006	0.004	0.002	0.004	0.004	0.004											
<i>R. rubropilosum</i>	0.057	0.048	0.048	0.048	0.052	0.006	0.004	0.002	0.004	0.004	0.004	0.004										
<i>R. sinii</i>	0.054	0.046	0.046	0.046	0.050	0.006	0.004	0.002	0.004	0.004	0.004	0.004	0.006									
<i>R. taiwanalpinum</i>	0.046	0.041	0.041	0.041	0.046	0.039	0.037	0.034	0.037	0.037	0.037	0.037	0.036	0.002								
<i>R. mariesii</i>	0.048	0.041	0.041	0.041	0.046	0.045	0.043	0.041	0.043	0.048	0.043	0.043	0.045	0.043	0.034							
<i>R. lamprophyllum</i>	0.052	0.046	0.046	0.046	0.050	0.050	0.048	0.045	0.048	0.052	0.048	0.050	0.050	0.048	0.041	0.043	0.006					
<i>R. ovatum</i>	0.048	0.043	0.043	0.043	0.048	0.059	0.057	0.055	0.057	0.062	0.057	0.059	0.059	0.057	0.045	0.050	0.043	0.048				
<i>R. ellipticum</i>	0.043	0.039	0.039	0.039	0.043	0.052	0.050	0.048	0.050	0.054	0.050	0.052	0.052	0.050	0.048	0.045	0.043	0.047	0.050			
<i>R. kawakamii</i>	0.193	0.196	0.196	0.196	0.201	0.195	0.193	0.196	0.193	0.198	0.193	0.196	0.195	0.192	0.190	0.195	0.201	0.204	0.050			
<i>G. itoana</i>																						0.203

cluster III. In addition, *R. mariesii*, *R. ellipticum*, and *R. kawakamii* formed three independent clusters (Figure 3). Based on the systematics of *Rhododendron* (Yang *et al.*, 1999; He *et al.*, 1999), these 20 *Rhododendron* species in Taiwan were classified into four subgenera, of which *R. kawakamii* belongs to the subgenus *Rhododendron*; *R. lamprophyllum*, *R. ovatum*, and *R. ellipticum* belong to the subgenus *Azaleastrum*; *R. hyperythrum*, *R. morri*, *R. formosanum*, *R. pseudo-chrysanthum*, and *R. rubropunctatum* belong to the subgenus *Hymenanthes*; and *R. lasiostylum*, *R. rubropilosum*, *R. kanehirai*, *R. breviperulatum*, *R. oldhamii*, *R. noriakianum*, *R. nakaharai*, *R. taiwanalpinum*, *R. longiperulatum*, *R. sinii*, and *R. mariesii* belong to the subgenus *Tsutsusi*. *Rhododendron ellipticum* is separated from the other species and is closer to cluster II (including *R. lamprophyllum* and *R. ovatum*). This result supports *R. ellipticum* (section *Choniastrum*) and cluster II species (section *Azaleastrum*) all being under the subgenus *Azaleastrum*. In addition, *R. mariesii* (section *Brachycalyx*) is another unique species and more closely aligned to cluster I (section *Tsutsusi*). Both of these groups are under the subgenus *Tsutsusi* (Table I and Figure 3). Therefore, molecular data from ITS sequences in this study are in agreement with the traditional systematics of *Rhododendron* in Taiwan.

According to revisions of the genus *Rhododendron* in Taiwan described by Lu and Yang (1989) and Li *et al.* (1998), *R. hyperythrum*, *R. morri*, *R. rubropunctatum*, and *R. pseudo-chrysanthum* were combined into one species; *R. longiperulatum*, *R. sinii*, and *R. nakaharai* were combined into one species; and *R. breviperulatum* and *R. lasiostylum* were combined into one species. Basically, the findings of ITS sequence analysis support those revisions of the genus *Rhododendron* in Taiwan by Lu and Yang (1989) and Li *et al.* (1998). Molecular data from the *trnF-trnL* gene sequence of cpDNA also showed that *R. hyperythrum*, *R. morri*, *R. rubropunctatum*, and *R. pseudo-chrysanthum* should be combined into one species (Hwang and Hsu, 2001). Furthermore, from the phylogenetic tree, the genetic relationships among the ten *Rhododendron* species in cluster I (members of the section *Tsutsusi*) are very close, although these ten species of the section *Tsutsusi* in Taiwan still have questions involving their classification. Therefore, more species of the section *Tsutsusi* from other localities and more molecular data should be analysed to evaluate the genetic relationship within the section *Tsutsusi* in the future.

In conclusion, the genetic relationships of the 20 *Rhododendron* species in Taiwan could be established based on the sequence of the ITS of rDNA. The findings in this study agree with the traditional systematics of *Rhododendron* in Taiwan. In the future, the ITS sequence may be applied to evaluate the systematics of *Rhododendron* species in the world. Furthermore, this study also revealed that members of the section *Tsutsusi* of *Rhododendron* in Taiwan still have some questions at the species level.

We are grateful to Dr T. Y. Chou for his valuable comments and helpful discussions in the course of the study.

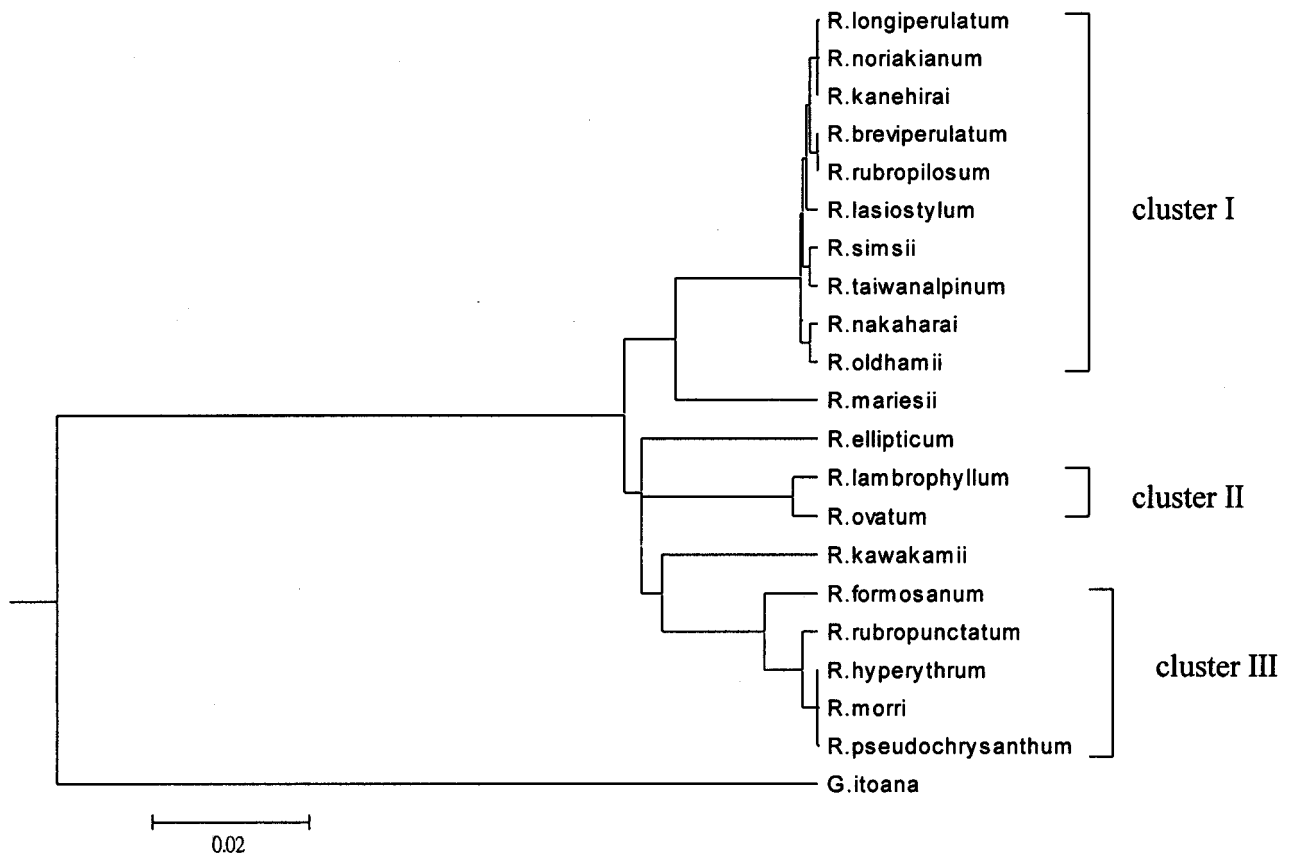


FIG. 3

A dendrogram of the 20 species of the genus *Rhododendron* in Taiwan obtained from sequence comparison of the ITS region of rDNA.

REFERENCES

- BALDWIN, B. G. (1993). Molecular phylogenetics of *Calycadenia* (Compositae) based on ITS sequences of nuclear ribosomal DNA: chromosomal and morphological evolution reexamined. *American Journal of Botany*, **80**, 222–38.
- BEAN, W. J. (1976). *Trees and shrubs hardy in the British Isles III (N-Rh)*, 8th ed. John Murray, London.
- CHASE, M. W. and HILLIS, H. G. (1991). Silica gel: an ideal material for field preservation of leaf samples for DNA studies. *Taxon*, **40**, 215–20.
- DOYLE, J. and DOYLE, J. (1987). A rapid DNA isolation procedure for small quantities of fresh leaf tissue. *Phytochemical Bulletin*, **19**, 11–5.
- HE, M. Y., FANG, M. Y., HU, W. K. and HU, L. C. (1999). Ericaceae (2). In: *Angiospermae, Dicotyledoneae*. (Hu, L. C. and Fang, M. Y., Eds). Flora Republicae Popularis Sinica. Tomus **57**(2). Science Press, Beijing. (in Chinese).
- HWANG, S. Y. and HSU, K. K. (2001). Molecular phylogeny of eight Taiwanese *Rhododendron* species on chloroplast *trnF-trnL* DNA sequences. *Taiwan Journal of Forest Science*, **16**, 153–60. (in Chinese)
- IQBAL, J. M., PADEN, D. W. and RAYBURN, A. L. (1995). Clonal stability of RAPD markers in three *Rhododendron* species. *Journal of Experimental Horticulture*, **13**, 43–6.
- KIMURA, M. (1980). A simple method for estimating evolutionary rates of base substitution through comparative studies of nucleotide sequences. *Journal of Molecular Evolution*, **16**, 111–20.
- KISS, T., SZKUKALEK, A. and SOLYMOSEY, F. (1989a). Nucleotide sequence of a 17S (18S) rRNA gene from tomato. *Nucleic Acids Research*, **17**, 2127.
- KISS, T., KIS, M. and SOLYMOSEY, F. (1989b). Nucleotide sequence of a 25S rRNA gene from tomato. *Nucleic Acids Research*, **17**, 796.
- KRON, K. A. (1996). Phylogenetic relationships of Empetraceae, Epacridaceae, Ericaceae, Monotropaceae, and Pyrolaceae: evidence from nuclear ribosomal 18S sequence data. *Annals of Botany*, **77**, 293–303.
- KRON, K. A. and CHASE, M. W. (1993). Systematics of Ericaceae, Empetraceae, Epacridaceae, and related taxa upon *rbcL* sequence data. *Annals of Missouri Botanical Garden*, **80**, 735–41.
- KRON, K. A., GAWEN, L. M. and CHASE, M. W. (1993). Evidence for introgression in azaleas (*Rhododendron*; Ericaceae): chloroplast DNA and morphological variation in a hybrid swarm on Stone Mountain, Georgia. *American Journal of Botany*, **80**, 1095–99.
- KRON, K. A. and JUDD, W. S. (1990). Phylogenetic relationships within the Rhodora (Ericaceae) with specific comments on the placement of *Ledum*. *Systematic Botany*, **15**, 57–68.
- KRON, K. A. and JUDD, W. S. (1997). Systematics of the *Lyonia* group (Andromedeae, Ericaceae) and the use of species as terminals in higher-level cladistic analyses. *Systematic Botany*, **22**, 479–92.
- KRON, K. A., JUDD, W. S. and CRAYN, D. M. (1999). Phylogenetic analysis of Andromedeae (Ericaceae subfam. Vaccinioideae). *American Journal of Botany*, **86**, 1290–300.
- KUMAR, S., TAMURA, K., JAKOBSEN, I. B. and NEI, M. (2001). MEGA 2.1: Molecular Evolutionary Genetics Analysis software. Arizona State University, Tempe, USA.
- LI, H. L. (1978). Ericaceae. In: *Flora of Taiwan*, **4**. (Li, H. L. et al., Eds). Epoch Publishing Co., Taipei, Taiwan, 21–38.
- LI, H. L., LU, S. Y., YANG, Y. P. and TSENG, Y. H. (1998). Ericaceae. In: *Flora of Taiwan*, 2nd ed., Vol. **4**. Editorial Committee of the Flora of Taiwan, Second Edition, Taipei, Taiwan, 17–39.
- LU, S. Y. and YANG, Y. P. (1989). A revision of *Rhododendron* (Ericaceae) of Taiwan. *Bull. Taiwan Forest Research Institute New Series*, **4**, 155–66. (in Chinese, with English summary)
- RAYBURN, L. A., IQBAL, M. J. and PADEN, D. W. (1993). Positive identification of *Rhododendron* through DNA fingerprinting. *Journal of the American Rhododendron Society*, **47**, 137–8.
- RITLAND, C. and STRAUS, N. A. (1993). High evolutionary divergence of the 5.8S ribosomal DNA in *Mimulus glaucescens* (Scrophulariaceae). *Plant Molecular Biology*, **22**, 691–6.

- ROHLF, F. J., KISHPAUGH, J. and KIRK, D. (1982). *Numerical taxonomy system of multivariate statistical programs*. State Univ. New York, Stony Brook, NY.
- SANG, T., CRAWFORD, D. J. and STUESSY, T. F. (1995). Documentation of reticulate evolution in peonies (*Paeonia*) using internal transcribed spacer sequences of nuclear ribosomal DNA: implications for biogeography and concerted evolution. *Proceedings of the National Academy of Science USA*, **92**, 6813–7.
- SCHEIBER, S. M., JARRET, R. L., ROBACKER, C. D. and NEWMAN, M. (2000). Genetic relationships within *Rhododendron* L. section *Pentanthera* G. Don based on sequences of the internal transcribed space (ITS) region. *Scientia Horticulturae*, **85**, 123–35.
- SLEUMER, H. (1966). Ericaceae. In: *Flora Malesiana Series I*, **6**, 470–668.
- SUN, Y., SKINNER, D. Z., LIANG, G. H. and HULBERT, S. H. (1994). Phylogenetic analysis of sorghum and related taxa using internal transcribed spacers of nuclear ribosomal DNA. *Theoretical and Applied Genetics*, **89**, 26–32.
- TAKAIWA, F., OONO, K. and SUGIURA, M. (1984). The complete nucleotide sequence of a rice 17S rRNA gene. *Nucleic Acids Research*, **12**, 5441–8.
- TAKAIWA, F., OONO, K. and SUGIURA, M. (1985). The complete nucleotide sequence of a rice 25S rRNA gene. *Gene*, **37**, 255–89.
- WILLIS, J. C. (1985). *A dictionary of the flowering plants and ferns*. Cambridge University Press, UK.
- YANG, H. B., FANG, R. C. and CHIN, T. L. (1999). Ericaceae (1). In: *Angiospermae, Dicotyledoneae* (Fang, R. C., Ed.). *Flora Reipublicae Popularis Sinica*. Tomus **57**(1). Science Press, Beijing. (in Chinese).